Reappraisal and neotypification of Phyllachora feijoae

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Abstract: Acca sellowiana (Myrtaceae), feijoa (in Brazil, goiaba da serra), is a native southern South America tree that produces edible fruits which, although only occasionally cultivated in South America, became a significant fruit crop in New Zealand. Recently, during surveys for fungal pathogens of feijoa in southern Brazil, several plants were found bearing tar-spot symptoms caused by a species of *Phyllachora*. A literature search enabled us to identify the fungus as *Phyllachora feijoae*, a little-known species originally described in the 19th century by H. Rehm and later transferred to the genus *Catacauma*. The name *Catacauma feijoae*, although now regarded as a later synonym of *P. feijoae* is still mistakenly in use (as, for instance, in the Brazilian list of fungi on plants). The type specimen was most probably deposited in the Botanisches Garten und Museum Berlin-Dahlem (B) and lost or destroyed during World War II, and could not be located. The recent recollection of abundant material of this fungus in the vicinity of Pelotas (Rio Grande do Sul, Brazil) allowed its re-examination and neotypification. *Phyllachora feijoae* is also illustrated here for the first time.

Key words:

Ascomycota
Brazil
fruit crop
Myrtaceae
Neotropics
nomenclature
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INTRODUCTION

The plant family Myrtaceae includes approximately 150 genera with over 5 500 species (Heywood et al. 2007), amongst which are some important forestry species (e.g. Eucalyptus spp.) and several fruit crops such as guava (Psidium guajava). Some, such as Acca sellowiana (common name feijoa; in Brazil, goiabeira da serra) are only minor fruit crops. Acca sellowiana is a shrub or small tree native to southern South America (southern Argentina, Brazil, Paraguay, and Uruguay) and, although only occasionally cultivated in South America, it has become more significant as a fruit crop in New Zealand (Al-Harthy 2010). There are few published records of fungal pathogens associated with feijoa (Farr & Rossman 2011, Mendes & Urben 2011). However, during a recent search for pathogens of feijoa in the southern Brazilian state of Rio Grande do Sul, individuals of A. sellowiana in rural areas in the vicinity of Pelotas had foliage with intense tar-spot symptoms. Such symptoms were typical of those caused by fungi belonging to the genus Phyllachora. Examination of specimens collected and a literature and herbarium search were performed in order to clarify the identity of the fungus on feijoa, and the results of these investigations are presented here.

MATERIAL AND METHODS

Samples of diseased foliage of *Acca sellowiana* were collected in two localities. These were dried in a plant press and taken

to the laboratory for further examination. Representative specimens were deposited in the local herbarium (Herbarium Universidade Federal de Viçosa, VIC). Examination of selected leaves bearing tar-spot symptoms with the help of an Olympus SXZ7 stereoscopic microscope revealed that fungal structures were immersed in the leaf tissue and sections were prepared and mounted in lactophenol and lactofucsin for further examination. Additionally, sections were also prepared with a freezing microtome (Cryostat Microm® HM 520). Observations, photographs, and line drawings were prepared with a light microscope Olympus BX51, fitted with a digital camera (Olympus E-volt 330) and a drawing tube.

TAXONOMY

Phyllachora feijoae Rehm, Hedwigia **36**: 370 (1897). Synonym: Catacauma feijoae (Rehm) Theiss & Syd., Ann. Mycol. **13**: 397 (1915). (Fig. 1)

Type: Brazil: Rio Grande do Sul: Pelotas, Chácara da Brigada, Cerro da Buena, on leaves of Acca sellowiana (Myrtaceae), 18 Aug. 2010, R. W. Barreto (VIC 31476 – neotype designated here; B 70 0015054 – isoneotype).

Other specimen examined: **Brazil**: Rio Grande do Sul: Pelotas, Capão do Leão, on leaves of Acca sellowiana (Myrtaceae), 18 Aug. 2010, R. W. Barreto (VIC 31766).

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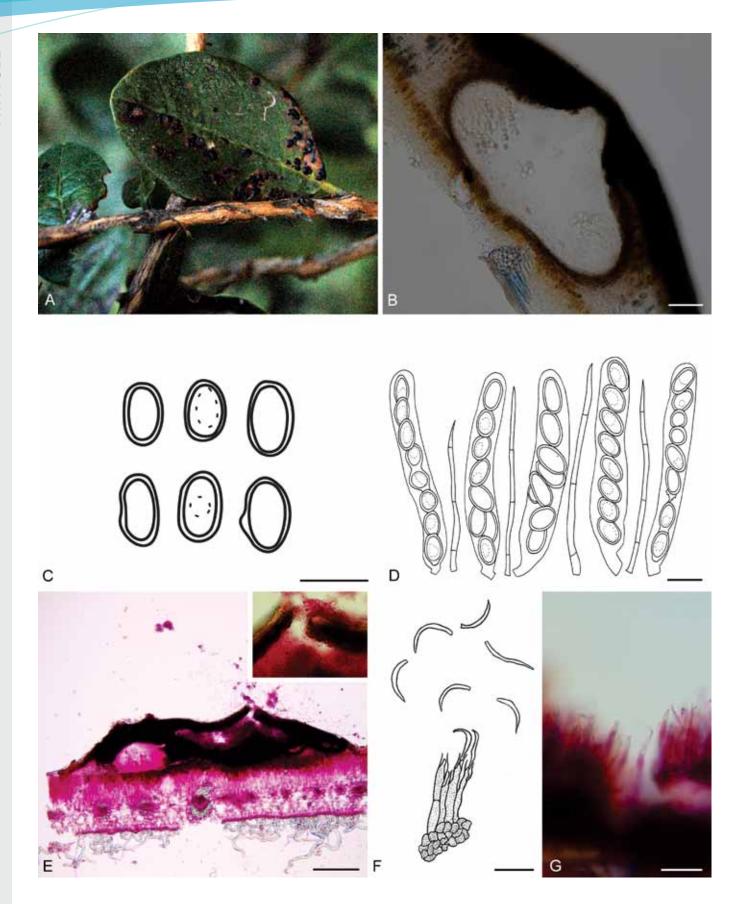


Fig. 1. Phyllachora feijoae (VIC 31766). **A.** Tar-spots on leaves of *Acca sellowiana*. **B.** Perithecium and clypeus. **C.** Ascospores. **D.** Paraphyses and asci with ascospores. **E.** Ascomata (*left*) and conidioma (*right*) and close up part of the conidioma. **F–G.** Conidiogenous cell and conidia. Bars: $B = 80 \mu m$, $C = 20 \mu m$, $D = 20 \mu m$, $E = 275 \mu m$, $E = 275 \mu m$.

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Lesions on living leaves, adaxially on all leaves at various developmental stages, initially punctiform, becoming irregular tar-spots, raised, with age surrounded by yellowish to reddish peripheral necrotic haloes, widely distributed and leading to foliage distortions, 0.2-0.3 × 2.1-3.0 mm diam, indistinct abaxially. Internal mycelium intra- and intercellular, hyphae $2.0\mbox{--}3.0~\mu\mbox{m}$ diam, branched, septate, hyaline to pale brown. External mycelium absent. Stromata adaxial, clypeate, shieldlike, merged with the upper wall of the ascoma. Conidia formed within stromata externally indistinguishable from teleomorph stomata; flattened, lenticular to irregular (in section), epigenous, subepidermal, single or in combination with ascomata, sometimes very broad occupying nearly the whole breadth of the stroma, 615–1729 x 100–184 µm walls of dark brown textura angularis, 38.5-69 µm thick, smooth; conidiogenous cells subcylindrical, straight, 15–25(–40) x 2–3 µm, 0–1-septate, pale brown; conidia mucilaginous, enteroblastic, acicular, curved, lunate or sigmoid, 13–19 x 1.5 µm, aseptate, thin-walled, hyaline, smooth. Ascomata perithecial, epigenous, immersed, solitary, spherical to subsphaerical, somewhat to strongly depressed, short papillate, 41-218 µm diam, inconspicuously ostiolate, composed of thin-walled brown textura angularis, walls 6.5-44 μm, 7–11 cells thick, outer layers dark brown, inner layers pale brown to subhyaline. Interascal tissue of paraphyses, 2.5-3 µm diam, longer than the asci, filiform, septate, hyaline, thinwalled, constricted at the septae; periphyses well-developed, filiform, hyaline, thin-walled. Asci unitunicate, cylindrical to clavate, short-stalked, 70.5-104 x 13-27 µm, apex broadly rounded to nearly flat, thin-walled, 8-spored. Ascospores at first uniseriate but sometimes partially biseriate, 15.5–22 x 8–14 µm, ellipsoidal to cylindric-ellipsoidal, rounded at the ends, walls 2-3 µm thick, aseptate, hyaline, smooth, without a mucous sheath or appendages.

Notes: Very little information is available on Phyllachora feijoae. Only a very brief description is given in the original publication of Rehm (1897). Later, Theissen & Sydow (1915) prepared a more complete description of the fungus when combining it into Catacauma. This is, nevertheless, somewhat incomplete and no illustrations were provided. Furthermore, the description was apparently based on Rehm's material collected in "Serra Geral, Minas Gerais - Brazil". The last publication dealing with this fungus was that of Jimenez & Hanlin (1992), where names of fungi described in Catacauma were listed. Although the authors acknowledged that after Petrak's (1924) work it became widely accepted that the distinction of Catacauma from Phyllachora was artificial, they prudently did not propose that names in Catacauma should be immediately rejected or recombined into Phyllachora without a careful re-examination of types. Since that publication, mycologists have shown little interest in the names of fungi referred to Catacauma, but some earlier fungal names in Phyllachora have been reinstated. That is the case of the name C. feijoae, presently listed in MycoBank and Index Fungorum as a later synonym of P. feijoae. Nevertheless, this name is still being used in other instances (e.g. the Brazilian list of fungi on plants; Mendes & Urben 2011).

An expanded description based on the material recently collected in Brazil is provided above. This is also the first time illustrations of *P. feijoae* have been published. The original material of the species studied by Rehm would almost certainly have been deposited in the collections of the Botanisches Garten und Museum Berlin-Dahlem (B), but if so it appears to have been lost or destroyed during World War II as it could not now be found (H.J.M. Sipman, pers. comm.). We therefore designate one of the recent collections as a neotype to fix the application of the name.

DISCUSSION

The fungus on Acca sellowiana exhibits all the typical features, both in terms of symptoms produced on the host and in its morphology, to members of the genus Phyllachora (Phyllachoraceae, Phyllachorales). Phyllachora is a large genus including approx. 1000 named species (Kirk et al. 2008). All species of Phyllachora are biotrophic plant pathogens, causing tar-spots on members of numerous plant families, but are particularly common on Fabaceae (Cannon 1991) and Poaceae (Parbery 1971). Besides the presence of a well-developed, dark brown to black clypeus, other features such as the formation of the perithecia within the plant tissues, and hyaline, thin-walled, smooth and aseptate ascospores, are typical for the genus (Cannon 1991). Around 70 species of Phyllachora have been described on members of Myrtaceae worldwide (Farr & Rossman 2011), with 21 species recorded on this host-family in Brazil (Mendes & Urben 2011). Species of Phyllachora associated with Myrtaceae have never been monographed.

Some of the older records of Phyllachora on Myrtaceae were later recognized as mistakenly placed in that genus. Some were found to belong to other genera, - such as P. pululahuensis (now regarded as a synonym of Vestegrenia multipunctata; von Arx & Müller 1954), and P. eucalypti (now recognized as a synonym of Clypeophysalospora latitans; Crous et al. 1990). Other species were recombined into genera such as P. peribebuyensis which is now treated as Coccodiella peribebuyensis (Katumoto 1968). Several names in Phyllachora that are listed on members of Myrtaceae were found to be later synonyms of already known species names: P. conspurcata (syn. P. tropicalis; Saccardo 1883), P. phylloplaca (syn. P ipirangae; Theissen & Sydow 1915b), P. pseudostromatica (syn. P. melaleuca; Sydow & Sydow 1904), and P. semillunata (syn. P. selenospora; Petrak & Ciferri 1930). Additionally, P. langdonii is now treated as a subspecies of P. callistemonis, P. callistemonis subsp. langdonii (Pearce & Hyde 1994).

In the case of species of *Phyllachora* recorded from Brazil, an issue to be taken into consideration is that numerous species names are included in Mendes *et al.* (1998), and have also been kept in the database of fungi on plants in Brazil (Mendes & Urben 2011) but quoted as being "in press". These names, for which Medeiros & Dianese are given as authors, have never been validly published and include the following

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 Table 1. Data on Phyllachora spp. described on hosts belonging to Myrtaceae.

Species	Asci (µm)	Ascospores (µm)	Host plants	References
P. ambígua	50–60 x 8–12	9–11 x 6	Syzygium cumini (syn. Eugenia jambolana)	Theissen & Sydow (1915b)
P. angustispora	80-90 x 12-14	30 x 8–9	Eugenia sp.	Saccardo (1916)
P. bella	60-70 x 5-7	7.5–9 x 3–4	Syzygium australe (syn. E. australis)	Sydow (1937)
P. biareolata	90-95 x 6-9	12 x 5	Eugenia rhombea	Saccardo (1891)
P. biguttulata	50-65 x 8-10	10-12 x 5-5.5	Campomanesia rhombea	Saccardo (1913)
P. brenesii	70–80 x 10–15	12-17 x 8-10	Eugenia guayaquilensis	Sydow & Petrak (1929)
P. callistemonis	115–210 x 12.5–16	18–27.5 x 7.5–10	Callistemon pallidus	Pearce & Hyde (1994)
P. callistemonis subsp. Langdonii	100-154 x 12-20	18–25 x 6–9	Callistemon sp.	Pearce & Hyde (1994)
P. callistemonis subsp. Similis	117–173 x 18–27.5	18–29 x 7.5–12.5	Callistemon viminalis	Pearce & Hyde (1994)
P. capensis	100–120 x 13–14	60–70 x 5–6	Eugenia zuluensis	Doidge (1942)
P. cayennensis	68–75 x 12–14	20–24 x 7–8	Psidium sp.	Theissen & Sydow (1915b)
P. clavata	110-140 x 15-18	39–45 x 3–6	Myrcia sp.	Garces Orejuela (1944)
P. curvulispora	60-80 x 10-20	17–20 x 5–7	Myrtaceae sp.	Saccardo (1925-1928)
P. distinguenda	60–70 x 18	18–20 x 4.5	Myrtaceae sp.	Saccardo (1899)
P. egenula	70–85 x 7–8	10–13 x 5-6	Leptospermum lanigerum	Sydow (1938)
P. emarginata	80–130 x 18–28	16–20 x 10–12	Eugenia sp.	Petrak (1948)
P. eugeniae	60-75 x 7-9	8-10 x 4-4.5	Eugenia rhombea	Chardón (1927)
P. feijoae	60 x 25	18 x 10	Acca sellowiana	Rehm (1915)
P. gentilis	120 x 4–12	18–20 x 8–9	Eugenia sp.	Saccardo (1895)
P. goyazensis	70–90 x 17–18	12-14 x 8-12	Myrtaceae sp.	Hennings (1895)
P. guavira	100-110 x 6-8	12 x 5	Psidium sp.	Theissen & Sydow (1915b)
P. ipirangae	30-90 x 10-12	15–16 x 8	Eugenia sp.	Theissen & Sydow (1915b)
P. lindmanii	80–90 x 13–16	16-24 x 13-16	Myrtaceae sp.	Theissen & Sydow (1915a)
P. maculata [*]		22–25	Eucalyptus sp.	Cooke (1891)
P. manuka		10.5–13 × 6.5–8	Leptospermum scoparium	Johnston & Cannon (2004)
P. melaleucae	66-84 x 8-11		Melaleuca spinosa	Theissen & Sydow (1915a)
P. myrciae*			Eugenia bimarginata	Saccardo (1883)
P. myrciae-rostratae	100-120 x 6-8	14–17 x 5–6	Myrcia splendens (syn. M. rostrata)	Viégas (1944)
P. muelleri	95–120 x 13–15	28–32 x 6–7	Eugenia dodonaeifolia	Chardón <i>et al.</i> (1940)
P. myrrhinii	50–72 x 12–16	14–16 x 5	Myrrhinium atropurpureum var. octandrum	Theissen & Sydow (1915a)
P. nigerrimum	100–130 x 9	10–16 x 5	Campomanesea adamantium (syn. C. caerulea)	Viégas (1944)
P. opaca	80–85 x 6–8	10 x 4-4.5	Myrtaceae sp.	Berlese & Voglino (1886)
P. peglerae	120–140 x 17–20	20-23 x 12-13	Eugenia capensis	Doidge (1942)
P. pettimenginii	85–105 x 14–18	2-8-32 x 8.5-11	Myrtaceae sp.	Maire (1908)
P. rhytismoides		14–19.5 × 12–15.5	Melaleuca cajuputi	Cannon (1991)
P. rickiana	68–78 x 14–15	10–13 x 6	Myrtaceae sp.	Theissen (1918)
P. rimulosa	85–100 x 10	14 x 8	Eugenia sp.	Saccardo (1925–1928)
P. samanensis	70–83 x 13–16.5	32-40 x 6-7.5	Eugenia sp.	Petrak & Ciferri (1932)
P. shivasii	136–225 x 10–15	15–22 x 6–8.5	Melaleuca viridiflora	Pearce & Hyde (1995)
P. subcircinans	80–90 x 10–16	14–16 x 8–10	Psidium grandifolium	Viégas (1944)
P. subopaca	75 x 10–15	12–14 x 7	Myrtaceae sp.	Saccardo (1899)
P. tachirensis	109–166 x 9.5–12	13–17 x 7–8	Eugenia sp.	Chardón & Toro (1934)
P. tropicalis	70–75 x 10–14	15–18 x 7–8	Psidium grandifolium	Saccardo (1883)
P. truncatispora	70–90 x 16–24	22–26 x 7–8	Myrtaceae sp.	Viégas (1944)
P. urbaniana	70–90 x 16–18	14–15 x 6–8	Myrtaceae sp.	Saccardo (1899)
P. verrucosa	78–105 x 15–19	14–20 x 9–13	Melaleuca leucadendra	Arx & Müller (1954)

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Table 1. (Continued).

Species	Asci (µm)	Ascospores (µm)	Host plants	References
P. whetzelii	87–109 x 8–10.5	11.5–13 x 3–4	Eugenia sp.	Chardón (1921)
P. woodiana	80-100 x 6-7.5	12.5–15.0 x 5–6	Eugenia capensis	Doidge (1942)

species designations associated with members of *Myrtaceae*: *P. eugenii-complicatae*, *P. eugenii-punctifolie*, *P. myrciae-decrescentis*, *P. myrciae-guianensis*, *P. myrciae-multiflorae*, *P. myrciae-multiflorae*, *P. myrciae-pallescentis*, *P. myrciae-tematae*, *P. myrciae-tortae*, and *P. pampulhae*. Although all these designations are not validly published, most may well represent good taxonomic species which are still awaiting formal description. Most were collected in the Brazilian cerrado, an area rich in endemic organisms of all kinds.

A study of the 48 published descriptions of taxa (including three varieties) of Phyllachora described from hosts belonging to Myrtaceae, is summarized in Table 1. This shows that there are three species of *Phyllachora* with close morphological similarity to *P. feijoae* on *A. sellowiana*: P. brenesii, P. emarginata, and P. subcircinans. Each of those species was found to have morphological differences from P. feijoae. Phyllachora. brenesii has perithecia with narrower walls (5 µm thick), and asci which are also narrower (10-15 µm wide). Phyllachora emarginata has thinner ascospore walls (2 µm). And P. subcircinans has much wider perithecia (250-500 µm diam). Additionally, P. feijoae can be recognized as distinct from the other species known on Myrtaceae (Table 1) by a combination of morphometric features; differences in perithecial diameter, ascus width, and the absence of a mucilaginous sheath on the ascospores. Although no comparison of the morphology of P. feijoae with other species on Myrtaceae was attempted in previous publications, our results indicate that this species is distinct from other Phyllachora species on this host-family, and so deserves recognition as a separate species. No significant discrepancies were found between the morphology of the neotype and the description provided in Theissen & Sydow (1915).

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